

Genome Sequence of the Bacteriocin-Producing Oral Probiotic *Streptococcus salivarius* Strain M18

Nicholas C. K. Heng, Nurul S. Haji-Ishak, Alaina Kalyan, Andrew Y. C. Wong, Marija Lovric, Joanna M. Bridson, Julia Artamonova, Jo-Ann L. Stanton, Philip A. Wescombe, Jeremy P. Burton, Mary P. Cullinan and John R. Tagg
J. Bacteriol. 2011, 193(22):6402. DOI: 10.1128/JB.06001-11.

Updated information and services can be found at:
<http://jb.asm.org/content/193/22/6402>

	<i>These include:</i>
REFERENCES	This article cites 13 articles, 5 of which can be accessed free at: http://jb.asm.org/content/193/22/6402#ref-list-1
CONTENT ALERTS	Receive: RSS Feeds, eTOCs, free email alerts (when new articles cite this article), more»

Information about commercial reprint orders: <http://jb.asm.org/site/misc/reprints.xhtml>
To subscribe to to another ASM Journal go to: <http://journals.asm.org/site/subscriptions/>

Genome Sequence of the Bacteriocin-Producing Oral Probiotic *Streptococcus salivarius* Strain M18

Nicholas C. K. Heng,^{1*} Nurul S. Haji-Ishak,¹ Alaina Kalyan,¹ Andrew Y. C. Wong,¹ Marija Lovrić,¹ Joanna M. Bridson,¹ Julia Artamonova,¹ Jo-Ann L. Stanton,² Philip A. Wescombe,³ Jeremy P. Burton,³ Mary P. Cullinan,¹ and John R. Tagg^{3,4}

Sir John Walsh Research Institute, Faculty of Dentistry,¹ and Department of Anatomy,² University of Otago, Dunedin, New Zealand; BLIS Technologies Ltd., Centre for Innovation, Dunedin, New Zealand³; and Department of Microbiology and Immunology, University of Otago, Dunedin, New Zealand⁴

Received 12 August 2011/Accepted 12 September 2011

***Streptococcus salivarius* is a Gram-positive bacterial commensal and pioneer colonizer of the human oral cavity. Many strains produce ribosomally synthesized proteinaceous antibiotics (bacteriocins), and some strains have been developed for use as oral probiotics. Here, we present the draft genome sequence of the bacteriocin-producing oral probiotic *S. salivarius* strain M18.**

The Gram-positive bacterium *Streptococcus salivarius* is a pioneer colonizer of the human oral cavity, and large populations persist at this site for the host's lifetime (12). *S. salivarius* is the prototype species of the *S. salivarius* group, which includes the important dairy species *Streptococcus thermophilus* (6). Many *S. salivarius* strains produce ribosomally synthesized proteinaceous antibiotics (bacteriocins; reviewed in reference 14), typically encoded by megaplasmid-borne loci (10). As *S. salivarius* is generally associated with good oral health, several bacteriocinogenic strains with proven safety records have been developed as oral probiotics (2–4, 8, 12).

S. salivarius M18 (formerly strain Mia) is a megaplasmid-carrying oral probiotic exhibiting broad-spectrum inhibitory activity against several streptococcal pathogens, notably the caries-causing *Streptococcus mutans* (10). In order to provide a genetic basis for factors enhancing its probiotic candidature, e.g., bacteriocin repertoire and colonization-related genes, and also to establish whether the strain is free of virulence factors and antibiotic resistance determinants, the *S. salivarius* M18 genome was sequenced by a whole-genome shotgun strategy using a Roche GS-FLX pyrosequencer (7). Approximately 42.9 million base pairs (~18-fold coverage) was assembled by Roche GS *de novo* assembler (versions 1.1.03.24 and 2.3) into ~150 contigs. All putative chromosomal contigs were ordered relative to the megaplasmid-free *S. salivarius* CCHSS3 genome sequence (GenBank accession number FR873481). Gap closures were achieved by direct Sanger-based sequencing of PCR amplicons generated with specific primers designed for contig termini.

The high-quality draft *S. salivarius* M18 chromosome sequence currently comprises five supercontigs (2,142,944 bp; GC content of 39.6%). The remaining genomic gaps contain multiple copies of large (>6-kb) genes encoding putative highly repetitive serine-rich proteins homologous to the *Strep-*

tococcus gordonii Hsa adhesin (9). The latter, which are conspicuously absent in *S. thermophilus*, may aid *S. salivarius* in colonizing oral surfaces. Automated annotation carried out by the rapid annotations using subsystems technology (RAST) (1) and NCBI Prokaryotic Genomes Automatic Annotation Pipeline (PGAAP) servers revealed 1,975 protein-coding sequences (CDSs), six rRNA operons, and 68 tRNA genes. A variety of insertion sequences were identified, with ISSag8 and IS1193 being the most common. In addition, the chromosome contains a locus (*slm*) specifying the production of a new anti-*S. mutans* lantibiotic bacteriocin designated salivaricin M.

The *S. salivarius* M18 megaplasmid, pSsal-M18, is 183,037 bp long (GC content of 34.8%) and is the first completely sequenced streptococcal megaplasmid. Of 172 CDSs annotated by PGAAP, 16 belong to the known bacteriocin-encoding loci for salivaricins A2 (11), 9 (13), and MPS (5, 14). Interestingly, there is a region of several open reading frames (ORFs) (flanked by insertion sequences) nearly identical to chromosomal genes found in *S. salivarius* strains CCHSS3 and 57.I (GenBank accession number CP002888), indicating gene exchange between the chromosome and megaplasmid. The *S. salivarius* M18 genome sequence will not only be useful for comparative genomics but is essential for the development of a functional genomics platform facilitating molecular evolution and ecological studies.

Nucleotide sequence accession numbers. This Whole Genome Shotgun project has been deposited at DDBJ/EMBL/GenBank under accession number AGBV00000000. The version described in this paper is the first version, AGBV01000000.

The M18 genome sequencing project was funded in part by grants from the Otago Medical Research Foundation (OMRF), the New Zealand Dental Association Research Foundation, and the New Zealand Lottery Grants Board. N.S.H.-I., A.K., A.Y.C.W., M.L., and J.M.B. were recipients of University of Otago Faculty of Dentistry Summer Research Studentships. J.A. was an OMRF Summer Research Scholar.

REFERENCES

1. Aziz, R. K., et al. 2008. The RAST Server: rapid annotations using subsystems technology. *BMC Genomics* 9:75.
2. Burton, J. P., et al. 2011. Evaluation of safety and human tolerance of the

* Corresponding author. Mailing address: Department of Oral Sciences, Faculty of Dentistry, University of Otago, P.O. Box 647, Dunedin 9054, New Zealand. Phone: 64 3 479-9254. Fax: 64 3 479-7078. E-mail: nicholas.heng@otago.ac.nz.

- oral probiotic *Streptococcus salivarius* K12: a randomized, placebo-controlled, double-blind study. *Food Chem. Toxicol.* **49**:2356–2364.
3. **Burton, J. P., et al.** 2010. Extended safety data for the oral cavity probiotic *Streptococcus salivarius* K12. *Probiotics Antimicrob. Prot.* **2**:135–144.
 4. **Burton, J. P., et al.** 2006. Safety assessment of the oral cavity probiotic *Streptococcus salivarius* K12. *Appl. Environ. Microbiol.* **72**:3050–3053.
 5. **Dodd, S. J.** 1999. A saliva model to demonstrate *Streptococcus salivarius* BLIS production and characterization of salivaricin MPS. M.S. thesis. University of Otago, Dunedin, New Zealand.
 6. **Facklam, R.** 2002. What happened to the streptococci: overview of taxonomic and nomenclature changes. *Clin. Microbiol. Rev.* **15**:613–630.
 7. **Heng, N. C. K., and J.-A. L. Stanton.** 2010. Oral bacterial genome sequencing using the high-throughput Roche Genome Sequencer FLX System. *Methods Mol. Biol.* **666**:197–218.
 8. **Tagg, J. R., and K. P. Dierksen.** 2003. Bacterial replacement therapy: adapting 'germ warfare' to infection prevention. *Trends Biotechnol.* **21**:217–223.
 9. **Takahashi, Y., et al.** 2002. Identification and characterization of *hsa*, the gene encoding the sialic acid-binding adhesin of *Streptococcus gordonii* DL1. *Infect. Immun.* **70**:1209–1218.
 10. **Wescombe, P. A., et al.** 2006. Megaplasmsids encode differing combinations of lantibiotics in *Streptococcus salivarius*. *Antonie Van Leeuwenhoek* **90**:269–280.
 11. **Wescombe, P. A., et al.** 2006. Production of the lantibiotic salivaricin A and its variants by oral streptococci and use of a specific induction assay to detect their presence in human saliva. *Appl. Environ. Microbiol.* **72**:1459–1466.
 12. **Wescombe, P. A., et al.** 2009. Streptococcal bacteriocins and the case for *Streptococcus salivarius* as model oral probiotics. *Future Microbiol.* **4**:819–835.
 13. **Wescombe, P. A., et al.** 2011. Salivaricin 9, a new lantibiotic produced by *Streptococcus salivarius*. *Microbiology* **157**:1290–1299.
 14. **Wescombe, P. A., N. C. K. Heng, J. P. Burton, and J. R. Tagg.** 2010. Something old and something new: an update on the amazing repertoire of bacteriocins produced by *Streptococcus salivarius*. *Probiotics Antimicrob. Prot.* **2**:37–45.